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Utilization of green formulation technique and efficacy estimation on cell line studies for dual anticancer drug therapy with niosomes

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Abstract

The aim of the present study was to prepare niosome formulations for the simultaneous encapsulation, dual drug therapy, of two anticancer drugs by the ecological probe sonication method. Poloxamer and sorbitan monostearate were used as surface active agents in niosomes, and the water soluble doxorubicin and poorly-water soluble paclitaxel were used as anticancer drugs. Thorough physicochemical analysis were performed for the niosomes, and their cytotoxicity and activity were evaluated on MCF-7 and PC3-MM2 cancer cell lines. Prepared niosomes were small in size with sizes ranging from 137 nm to 893 nm, and entrapment efficiencies were high, ranging from 91.24% to 99.99%. During the four weeks stability testing, the particle size remained stable. The niosomal formulations showed *in vitro* sustained drug release profiles for doxorubicin and clearly increased the dissolution rate of poorly water soluble paclitaxel. The incorporation of both the drugs into niosomes improved cell penetration and antiproliferative activity of the drugs PC3-MM2 cell lines. As a conclusion, doxorubicin and paclitaxel loaded niosome formulations resulted in relatively stable, small sized niosomes with improved drug release profiles, low toxicity, better cell penetration and antiproliferative activity. The niosomes showed synergistic effect due to the presence of both drugs, which can overcome multidrug resistance.

Keywords: antiproliferative activity, cell penetration, doxorubicin, dual drug therapy, paclitaxel, niosomes

1. Introduction

Cancer is one of the causative factors of death around the world in many countries (Yingchoncharoen *et al.*, 2016), and chemotherapy is among the approaches, which is effective against multiple cancers (Tahir *et al.*, 2017). However, the chemotherapeutics have toxic adverse effects on healthy living cells regardless of selectivity (Zheng *et al.*, 2015). Due to high toxicity and shorter half-life, the use of chemotherapeutics is limited to avoid toxicity. Accordingly, in order to avoid the hazardous adverse effects, the challenge is that chemotherapeutical agents are needed to be delivered only to the cancer cells with minimum delivery to normal cells (Yingchoncharoen *et al.*, 2016; Tahir *et al.*, 2017).

Different nanosized systems are the carrier of choice for the efficient loading of the drugs (hydrophilic or hydrophobic) in order to reach site specificity, prolonged circulation time in body, and lower toxicity (Sharma *et al.*, 2015). Niosomes, vesicular structures composed of non-ionic surfactants, are capable of encapsulating both hydrophobic as well as hydrophilic drugs (Sharma *et al.*, 2016). Niosomes have advantages over liposomes, like higher stability and entrapment efficiency, biocompatibility, non-immunogenicity and lower costs (Manconi *et al.*, 2002; Tavano *et al.*, 2014). A number of non-ionic surfactants, like alkyl ethers, alkyl esters, polysorbates, poloxamers and alkyl amides, have been used to produce niosomes (Di Marzio *et al.*, 2011; Escudero *et al.*, 2014; Moghassemi *et al.*, 2014).

Poloxamers are promising non-ionic polymeric surfactants, which have been used for niosome production and delivery of anticancer drugs for the treatment of multidrug resistant cancers. Pluronic L121 is one of the poloxamers used for the encapsulation of cytotoxic drugs, and it is also P-glycoprotein (P-gp) inhibitor (Yang *et al.*, 2007b). In most of the niosomal studies, only a single surfactant has been used (Di Marzio *et al.*, 2011;

Escudero *et al.*, 2014; Moghassemi *et al.*, 2014). However, when two or more non-ionic amphiphiles are used, more stable, small in size, monodispersed niosomes with better drug release profiles can be reached (Khan *et al.*, 2016).

Niosomes are produced by different methods, the most adopted being reverse phase evaporation, thin film hydration, ether injection and shaking methods (Kanaani *et al.*, 2017; Ravalika and Sailaja, 2017). These methods are time consuming and expensive, they use organic solvents, and after the production, solvent removal is laborious. Probe sonication method has been developed to overcome these problems (Dufes *et al.*, 2000). It is a green technique with low energy consumption and without the addition of any organic solvents. In this method, only aqueous drug phase is mixed with surfactants, cholesterol and other bilayer membrane additives (Khan *et al.*, 2017).

Doxorubicin (DOX) and paclitaxel (PXT), chemotherapeutic agents belonging to anthracycline and taxanes classes of cytotoxic drugs, respectively, are effective against number of cancers, including breast and prostatic cancers (Kim *et al.*, 2015; Pawar *et al.*, 2016). DOX is hydrophilic in nature, but, PXT is hydrophobic, which limits its bioavailability (Alemi *et al.*, 2018; Behnam *et al.*, 2018). Besides, the delivery of these cytotoxic drugs is challenging due to their highly toxic adverse effects and drug resistance (Liu *et al.*, 2017; Teixeira *et al.*, 2017; Sayed *et al.*, 2018). However, it has been shown that the combination of DOX and PXT have a great attraction due to synergistic effect with reduced systematic toxicity and higher antitumor efficacy (Devita *et al.*, 1975; Chabner and Roberts, 2005; Al-Lazikani *et al.*, 2012; Ag Seleci *et al.*, 2017; Alemi *et al.*, 2018, Yang *et al.*, 2019).

In the present study, co-delivery formulation of two anticancer drugs, DOX and PTX were prepared by an environmental friendly probe sonication method. The aims of the dual drug

therapy were synergistic effects with lower toxicity levels and higher antitumor efficacy. Niosomes containing only a single drug, as well as both the drugs, were formulated. Pluronic L121 and Span 60 surfactants were utilized for production of niosomes. Performance of niosomal formulations was confirmed by thorough physicochemical analysis and efficiency of the formulations were confirmed *in vitro* in different cancer cell lines.

2. Materials and methods

2.1 Materials

Doxorubicin hydrochloride (Fluorochem, UK) and paclitaxel (Fluorochem, UK) were studied chemotherapeutic drug substances. Sorbitan Monostearate (Span 60, Sigma-Aldrich, USA) and Polyethylene oxide - Polypropylene oxide - Polyethylene oxide (PEO-PPO-PEO) block copolymer (Pluronic L121, Mn 4400, Sigma-Aldrich, USA) were used as bilayer membrane formers in niosomes. Cholesterol (Sigma-Aldrich, USA) was used as membrane stabilizer and dicetylphosphate (DCP, Sigma-Aldrich, USA) as charge imparting agent. Hank's Balanced Salt Solution (HBSS), and Dulbecco's Modified Eagle's Medium (DMEM) were purchased from HyClone (USA). Tween 80 was used as a solubilizing agent in dissolution testing (Sigma-Aldrich, USA). Water used in all the tests was Milli-Q water (Millipore, Merckmillipore, USA).

2.2 Methods

2.2.1 Preparation of niosomes

The niosomes were prepared by probe sonication method (Khan *et al.*, 2019). First, the drugs, doxorubicin-HCl (DOX) and paclitaxel (PXT), individually or as a combination, were mixed with 15 mL of water with the aid of magnetic stirrer, after which cholesterol,

Span 60, Pluronic L121, and dicetylphosphate (DCP) were added. The compositions of different studied formulations are indicated in Table 1. The mixtures were then subjected to probe sonication (Vibra Cell, Sonics & Materials, Inc., USA) for 5 min time at 57°C probe temperature in a pulsatile manner (50 sec sonication with 10 sec pause) at an amplitude of 30%. After probe sonication, niosome formulations were collected and stored at 4°C for further characterization and cell line studies.

Table 1: Compositions of studied niosome formulations.

Formulations	Span 60 (mg)	Pluronic L121 (mg)	Cholesterol (mg)	DCP (mg)	DOX (mg)	PXT (mg)	Water (mL)
D1	43	290	77.3	1	2	-	15
D2	43	290	77.3	2	2	-	15
D3	43	290	77.3	0	2	-	15
D4	43	246	77.3	1	2	-	15
D5	43	334	77.3	1	2	-	15
P1	43	290	77.3	1	-	2	15
P2	43	290	77.3	2	-	2	15
P3	43	290	77.3	0	-	2	15
P4	43	246	77.3	1	-	2	15
P5	43	334	77.3	1	-	2	15
DP1	43	290	77.3	1	2	2	15
DP2	43	290	77.3	2	2	2	15
DP3	43	290	77.3	0	2	2	15
DP4	43	246	77.3	1	2	2	15
DP5	43	334	77.3	1	2	2	15
E1	43	290	77.3	1	-	-	15
E2	43	290	77.3	2	-	-	15
E3	43	290	77.3	0	-	-	15
E4	43	246	77.3	1	-	-	15
E5	43	334	77.3	1	-	-	15

2.2.2 Particle size, size deviation and zeta-potential

The diameter of the niosomes (z-average) and polydispersity index (PDI), based on dynamic light scattering (DLS) technique, as well as zeta-potential, were measured by using Zeta-sizer Nano ZS (Malvern Instruments Ltd., USA). The niosomal dispersions were diluted with water before the measurement to avoid multi scattering phenomenon. All the measurements were performed in triplicate.

2.2.3 Drug entrapment efficiency

For drug entrapment efficiency determinations, the formulations were ultracentrifuged at 14,500 rpm for 45 min time (Sigma Laborzentrifugen, D-37520, Germany). The supernatant was collected, the pellet at the bottom of the centrifuge tube was washed twice with water, water was collected, and centrifugation was repeated. Drug concentration in the aqueous portion of supernatants was determined. For PXT determination, high performance

liquid chromatography (HPLC, Agilent 1260, Agilent Technologies, USA), and for DOX, spectrophotometric analysis (Varioskan Flash, Thermo Fisher Scientific Inc., USA), were used. The percentage entrapment (EE%) of drugs were calculated according to the following equation (Equation 1) (Li *et al.*, 2016; Maestrelli *et al.*, 2017):

$$EE\% = [(Q_t - Q_r)/Q_t] \times 100, \quad (1)$$

where Q_t is the amount of drug initially used for the preparation of niosomes and Q_r is the amount of drug present in supernatant after centrifugation.

2.2.4 Transmission electron microscopy

The morphology of the niosomes was investigated by the transmission electron microscopy (TEM, Jeol JEM-1400, Jeol Ltd, Japan). For TEM analysis, small amount of niosomal dispersions were inserted on a carbon coated 200-mesh sized copper grid. The mesh was positioned horizontally for one minute, superfluous was removed with the aid of filter paper and one drop of 2% uranyl acetate was placed on the sample for staining (Somjid *et al.*, 2018).

2.2.5 Attenuated total reflectance - fourier transform infrared (ATR-FTIR) spectroscopy

The possible interactions between the drug, the non-ionic surfactants and other membrane additives were studied by attenuated total reflectance - fourier transform infrared (ATR-FTIR) spectroscopy. The ATR-FTIR spectra of all the individual components, their physical mixtures and niosome formulations containing DOX, PXT and DOX+PXT were measured. For ATR-FTIR analysis, the niosome dispersions were centrifuged, and the dried pellet was analyzed. The spectra were recorded by using FTIR spectrophotometer (Bruker Optics, Germany) with an additional horizontal accessory of ATR (MIRacle, Pike

Technology, Inc., Germany). The spectra were recorded at an ambient temperature between wavenumbers of 400-4500 cm^{-1} with 4 cm^{-1} resolution by using OPUS 5.5 software.

2.2.6 Thermal analysis

The physical states of the DOX and PXT in the formulations were estimated by using differential scanning calorimetry (DSC 823^e, Mettler Toledo, USA). Pure DOX, PXT, individual niosome constituents, their physical mixtures, and formulations containing DOX, PXT and DOX+PXT were accurately weighed (3-5 mg) in closed aluminum pan. For DSC analysis, the niosome dispersions were centrifuged, and the dried pellet was analyzed. The thermal scanning was carried out from 25°C to 260°C with a heating rate of 5°C/min. The analysis were conducted under the nitrogen gas flow (50 ml/min).

2.2.7 Stability studies

Four weeks stability study of niosome formulations was performed by storing the niosomal dispersions in sealed 20 mL glass vials at 4°C in refrigerator. The size, PDI and zeta-potential of the stored formulations were assessed at predefined time intervals (fresh samples, 1, 2, 3 and 4 weeks after production).

2.2.8 Dissolution studies

The dissolution studies were carried out in a glass vessel containing magnetic stirrer. HBSS with HEPES (Hanks' balanced salt solution with N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid) buffer pH 7.4 solution with 4% Tween 80 was used as dissolution medium. For the dissolution study, the aqueous dispersions of the formulations (2 mL) were put into the dissolution vessel containing dissolution medium. The study was conducted at 37°C, the amount of the medium was 250 mL, and stirring speed was 100 rpm. The aliquots were sampled at predefined time intervals (0, 15 min, 30 min, 45 min, 60 min, 1.5 h, 2 h, 3

h, 4 h, 6 h, 8 h, 12 h, and 24 h), and replaced with the same volume of fresh buffer. Samples withdrawn from the dissolution media were analyzed as such using a spectrophotometer with a wavelength of 480 nm for DOX, and HPLC for PXT.

Dissolution studies for the pure drugs in powdered form were carried out using the same protocol.

2.2.9 High performance liquid chromatography (HPLC)

In the HPLC method, the column used for the PXT detection was C18 ($4.6 \times 150 \text{ mm} \times 5 \text{ mm}$, Supelco Discovery C18, Phenomenex, USA), and the mobile phase used consist of water and acetonitrile (53:47, v/v). The flow rate was 1.0 mL/min, the temperature of column was 25°C, and the wavelength used for the drug detection was 227 nm. The injected volume of the drug solution was 20 μL .

2.2.10 Cell culturing

The MCF-7 breast cancer cells and PC3-MM2 human prostate cancer cells were grown in 75 cm² culture flasks (Corning Inc. Life Sciences, USA). The incubation was performed in 5% CO₂ in a gas incubator (Heraeus Instruments GmbH, Germany) at 37°C with 95% relative humidity. MCF-7 and PC3-MM2 cells were cultured in DMEM culture medium. The medium was supplemented with 1% non-essential amino acids (NEAA), 10% (v/v) fetal bovine serum (FBS), 1% L-glutamine, and 1% penicillin/streptomycin (PEST). Cells were thawed from the frozen stock and sub cultured at 80% confluency.

2.2.11 Cytotoxicity studies

The *in vitro* cytotoxicity of the niosomes was studied for the evaluation of the safety of the formulations. The viability of MCF-7 and PC3-MM2 cells was determined using an ATP-based cell viability kit.

Briefly, the cells were seeded in 96-well plates at a concentration of 2×10^5 cells/mL (100 μ L) and incubated overnight for the attachment. The niosomes were prepared in the medium with the concentration ranging from 25 to 1000 μ g/mL. The cell medium in 96-well plates was then replaced with the 100 μ L of the fresh medium containing niosomes and incubated for 24 h. After the incubation, the ATP-based viability was measured by further adding 100 μ L of reagent assay into each well (CellTiter-Glo Luminescent Cell Viability Assay, Promega, USA). The luminescence was measured with a Varioskan Flash plate reader (Thermo Fisher Scientific Inc., USA). The cells incubated with the cell culture medium and with Triton X-100 (1%) were measured as positive and negative controls, respectively. All the measurements were made triplicate.

2.2.12 Cell uptake studies

For qualitative evaluation of the cellular uptake of niosomes, 200 μ L of MCF-7 and PC3-MM2 cells were seeded into an 8-chamber slides (Nunc Lab-Tek II Chamber Slide System, Thermo scientific, Inc., USA) at a density of 2.5×10^4 cells per well, and incubated at 37°C overnight for proper attachment of the cells to the chamber. After the removal of the cell media, the cells were washed with HBSS-HEPES buffer (pH 7.4). Fluorescein isothiocyanate (FITC) labeled niosomes were prepared by loading the FITC during the aforementioned method for the preparation of drug loaded niosomes. 200 μ L of FITC labelled niosome suspension with different concentrations were added in each chamber and incubated at 37°C for 6 h time. After incubation, the cells were washed thrice with HBSS-HEPES buffer (pH 7.4) in order to remove the free niosomes and the cell membrane was stained with CellMask Deep Red (5 μ L/mL, Invitrogen, USA) for 3 min at 37°C. Cells were again washed with HBSS-HEPES buffer (pH 7.4), washed and fixed with 4% paraformaldehyde (PFA) for 15 min, and washed with HBSS-HEPES buffer (pH 7.4). After the nuclei was stained by adding 200 μ L of DAPI (4',6-diamidino-2-phenylindole, 2.8

$\mu\text{L}/\text{mL}$, Thermo Scientific, USA) for 5 min, cells were washed again and stored with 200 μL of HBSS-HEPES buffer (pH 7.4). The interaction of the niosomes with the cells was studied by a Leica SP5 inverted confocal microscope (Leica Microsystems, Germany), using a 63 \times 1.2-0.6 oil immersion objective.

2.2.13 Anti-proliferation assay

The *in vitro* cell growth inhibition of the drug-loaded niosomes was evaluated against the MCF-7 and PC3-MM2 cancer cells by cell proliferation experiments. The anti-proliferation effects of free DOX, PXT, and drugs loaded niosomal formulations, containing different concentrations of drugs (25-500 $\mu\text{g}/\text{mL}$) were measured using the previously described protocol for the cytotoxicity studies. All the experiments were repeated three times.

2.2.14 Statistical analysis

The statistical significance was determined using Student's t test (two-tailed); $P < 0.05$ was considered statistically significant in all the analyses (95% confidence level).

3. Results and Discussion

3.1 Preparation of niosomes

Niosomes were prepared with constant amounts of Span 60, cholesterol and drug materials (DOX and/or PXT), but with varying amounts of Pluronic L121 and DCP. In the niosome structure, DOX as hydrophilic drug is encapsulated inside the vesicles and PXT as hydrophobic drug is entrapped into the bilayer structure. Cholesterol is situated in the bilayer structure rigidifying it and minimizing drug leakage, and making the drug release controlled. Three different levels of the amount of Pluronic L121 and DCP were studied.

The central point composition for factorial design was selected to be 290 mg of Pluronic L121 and 1 mg of DCP. The exact compositions of all the niosome formulations are shown in Table 1.

3.2 Size and surface properties of niosomes

In this study, physicochemical parameters, size (<350 nm), PDI (<0.5) and zeta-potential (< -30 mV) were considered as critical quality attributes, CQAs, for the niosomes. The PDI value below 0.5 indicates low level of aggregation. Similarly, the zeta-potential value smaller than -30 mV means more stable niosomes with less aggregation tendency due to the electric repulsive forces (Mahale *et al.*, 2012). DCP was added to the composition in order to reach lower zeta-potential values. Previous studies have shown that niosomal formulations containing cholesterol are rigid, stable and intact, while niosomes without cholesterol forms gel like systems, and for that reason cholesterol was added to all the compositions (Somjid *et al.*, 2018).

The non-ionic amphiphiles having hydrophilic-lipophilic balance (HLB) value greater than 11, hinder the vesicular formation. Span 60, selected for this study, forms rigid, stable and large in size niosomes having high drug entrapment efficiencies due to larger particle size (Basiri *et al.*, 2017). Finally, the non-ionic polymeric surfactant, Pluronic L121, has the capability to entrap hydrophobic drugs, like PXT, efficiently inside the niosomes. And, it forms monodisperse and stable niosomes (Lee *et al.*, 2005; Lee *et al.*, 2011; Abdelbary and Tadros, 2013).

The negative charge-imparting agent in this study was DCP. It gives for particle population low polydispersity values, due to strong inter-niosomal repulsive forces induced by the high surface charge (Helal *et al.*, 2015). In this study, the size of all the niosomal formulations (with and without drug) were between 137 nm and 893 nm. The PDI ranged between 0.383

and 0.725 and zeta-potential values were from -26.5 mV to -49.1 mV (Table 2). Based on the size, size deviation and surface charge measurements, most of the studied niosomal formulations filled the above mentioned CQA requirements.

Table 2. Particle sizes, PDI values, zeta-potentials, and drug entrapment efficiency values for studied niosomal formulations.

Formulation	Size (nm)	PDI	Zeta-potential (mV)	%EE (DOX)	%EE (PXT)
D1	204.4 ± 8.7	0.470±0.069	-34.9± 0.5	92.90±0.13	-
D2	182.9 ± 20.7	0.423±0.009	-36.8±1.6	93.49±0.20	-
D3	241.1 ± 37.0	0.501±0.007	-34.3±2.6	92.32±0.18	-
D4	202.1 ± 18.0	0.510±0.033	-46.9 ±1.1	92.99±0.20	-
D5	190.1 ± 7.2	0.491±0.048	-39.6 ±1.1	92.73±0.20	-
P1	195.3 ± 21.5	0.462±0.055	-35.9± 2.0	-	99.96±0.00
P2	178.1 ± 3.0	0.417±0.027	-43.8± 0.6	-	99.98±0.00
P3	169.3 ± 19.2	0.435±0.098	-49.1± 1.3	-	99.98±0.00
P4	180.6 ± 7.2	0.447±0.076	-38.9±3.2	-	99.96±0.00
P5	173.5 ± 42.6	0.383±0.025	-47.0± 1.3	-	99.97±0.00
DP1	176.7 ± 11.4	0.493±0.010	-27.7±2.0	91.24±0.33	99.99±0.00
DP2	156.6 ± 10.2	0.450±0.009	-26.5± 0.7	92.58±0.39	99.99±0.00
DP3	147.6 ± 14.7	0.448±0.095	-32.1± 0.8	92.31±0.21	99.99±0.00
DP4	168.2 ± 11.8	0.481±0.072	-39.0 ±2.4	92.39±0.13	99.99±0.00
DP5	137.1 ± 4.1	0.437±0.046	-41.8±2.6	93.67±0.27	99.99±0.00
E1	195.6 ± 12.8	0.492±0.047	-27.5 ± 0.9	-	-
E2	236.3 ± 36.0	0.391±0.105	-27.5 ± 0.9	-	-
E3	443.5 ± 86.7	0.469±0.037	-34.9 ± 3.4	-	-
E4	300.5 ± 36.6	0.448±0.034	-38.8 ± 0.3	-	-
E5	893.6 ± 135.5	0.725±0.117	-39.9 ± 5.2	-	-

Three batches failed to fulfill the PDI criteria, but two of these (D3 and D4) were very close to the critical value (0.501 and 0.510). D3 formulation did not contain DCP, and D4 formulation contained the lowest amount of Pluronic L121 (246 mg). All the drug loaded niosomes were small in size, below 250 nm, but the size criteria was not fulfilled with two batches of empty niosomes E3 and E5 (E3 composition without DCP and E5 with the highest amount of Pluronic L121 (334 mg)). The zeta-potential value was not negative enough with four batches. However, the zeta-potential of these four batches was from -26.5 mV to -27.7 mV, which can still be considered high enough value to stabilize the niosomes. This was also confirmed in the stability studies, where all the batches showed good stability.

3.3 Drug entrapment efficiency (%EE)

The drug entrapment efficiency is very important property for drug formulations. In this study, the formulations that contained DOX, the %EE ranged between 92.32% and 93.49%.

The formulations containing only PXT, the %EE was even higher, being between 99.96% to 99.98%. With the formulations containing both DOX and PXT, entrapment efficiencies for DOX varied between 91.24% and 93.67%, while PXT had in all the formulations 99.99% entrapment efficiency value. Accordingly, the entrapment efficiencies were in very high level, especially entrapment of PXT was very close to 100%, and no real differences were seen in the entrapment efficiency values between different compositions with PXT.

The high entrapment efficiency of PXT was due to its very low water solubility. The affinity of the drug inside the niosomes is very high due to its lipophilicity: it has very high tendency to escape from the outer aqueous phase to niosomes. Aqueous solubility of DOX is higher and it can partly be left into the aqueous phase, shown by the slightly lower entrapment efficiency values. However, the drug entrapment efficiency values were generally high for both the drugs in all the tested compositions, and it can be concluded that the compositions of niosome formulations have very minor effect on the entrapment efficiencies.

In all the studied drug loaded formulations, the mean size of the niosomes was from 137 nm to 241 nm. The composition is affecting the particle size of the niosomes, and particle size can also affect the entrapment efficiency: it is typical that larger particles are able to entrap the drugs more efficiently. However, in this study the changes in particle sizes were that small that any conclusions of its impact on the drug entrapment could not be withdrawn.

Some examples of TEM images for niosomal formulations are shown in Figure 1. Based on the TEM analysis, the niosomes are close to spherical in shape and the sizes are corresponding well with the ones measured by DLS. No pure drug crystals were seen in TEM figures which is in accordance with the high drug entrapment values. This is also

supported by the fact that the PDI values of the empty niosomes were in the same level or even higher than corresponding values for drug loaded niosomes.

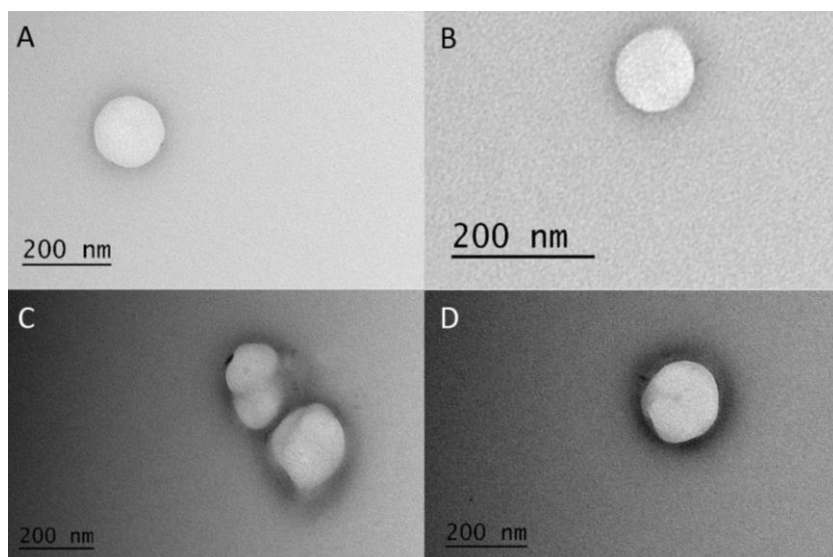


Figure 1. TEM images of E1 (figures A and B), and E2 (figures C and D) compositions of niosome formulations.

3.4 Stability studies

For stability testing purposes, all the niosomes were stored at 4°C for 4 weeks, and all the formulations were studied taking into account the above-mentioned CQAs as critical stability parameters. The niosomes (without drug) tend to increase in the size, but after one week in storage, they became stable. It is typical that after the process stress (here sonication), particle size can change a little bit before reaching a constant value. This is due to the relaxation process after the stress (sonication) phase is over. The particle sizes of the empty niosomes ranged between 195 nm and 893 nm with PDI values from 0.39 to 0.72, and zeta-potentials ranging from -27 mV to -39 mV as shown in Table 3. During the whole

storage time (4 weeks), all the drug-loaded niosomes were stable and only a little fluctuation in size was seen. Most stable formulations were the center point formulations D1, P1 and DP1 as well as formulations containing 2 mg of DCP, *e.g.* D2, P2 and DP2. The zeta-potential of the formulations D1, D2, P1, P2, DP1 and DP2 were deviating from the predetermined CQAs and was close to the limiting values. However, the low PDI values and stable particle sizes suggested a low level of aggregation and high stability of the niosomes, demonstrating good quality of these batches, too. The niosomes remained stable with low PDI values during the stability testing time as indicated in Table 3. The results indicated that higher quantities of DCP (1 or 2 mg) and medium quantity of Pluronic L121 (290 mg) showed the best stability of niosomes.

Table 3. Particle size, PDI and zeta-potential values of fresh (just after the production) and stored (at 4°C) niosomal formulations.

Time	Parameters	E1	E2	E3	E4	E5	D1	D2	D3	D4	D5
Fresh samples	Size (nm)	195.6±12.8	236.3±36.0	443.5±86.7	300.5±36.6	893.6±135.5	204.4±8.7	182.9±20.7	241.1±37.0	202.1±18.0	190.1±7.2
	PDI	0.492±0.047	0.391±0.105	0.469±0.037	0.448±0.034	0.725±0.117	0.470±0.069	0.423±0.009	0.501±0.007	0.510±0.033	0.491±0.048
	Zeta-potential (mV)	-27.5 ±0.9	-27.5± 0.9	-34.9± 3.4	-38.8±0.3	-39.9±5.2	- 34.9± 0.5	-36.8±1.6	- 34.3±2.6	- 46.9. ±1.1	- 39.6±1.1
1 week	Size (nm)	223.4±11.4	175.8±5.8	294.4±8.0	199.3±5.4	244.8±7.6	200.0±7.0	185.2±10.1	240.1±36.2	200.2±10.2	192.1±9.1
	PDI	0.340±0.046	0.198±0.040	0.362±0.038	0.282±0.014	0.535±0.020	0.341±0.011	0.314±0.071	0.430±0.017	0.490±0.033	0.451±0.011
	Zeta-potential (mV)	- 23.8±0.9	- 26.9± 0.9	- 28.2± 1.0	- 28.5 ±0.6	- 30.4±0.5	- 32.3± 0.1	- 34.0±1.5	- 34.1±2.2	- 46.1. ±2.2	- 36.8±2.1
2 weeks	Size (nm)	191.9±3.3	178.7±3.6	347.4±9.0	212.7±6.1	245.2±5.5	201.0±4.0	183.5±9.1	241.3±20.2	207.6±12.2	193.1±5.2
	PDI	0.168±0.039	0.169±0.028	0.375±0.030	0.299±0.022	0.543±0.015	0.340±0.019	0.310±0.011	0.415±0.011	0.450±0.011	0.410±0.034
	Zeta-potential (mV)	- 23.8±0.9	- 28.1± 2.2	- 29.5±2.5	- 28.2 ±3.0	- 26.3±0.5	- 32.5± 0.1	- 33.9±1.6	- 33.5±3.3	- 45.0 ±3.1	- 36.2±3.1
3 weeks	Size (nm)	199.4±4.9	172.2±1.7	296.3±12.6	208.7±4.6	291.6±8.3	203.1±2.6	182.9±11.2	253.6±26.4	215.2±12.2	201.3±5.1
	PDI	0.213±0.011	0.151±0.033	0.320±0.051	0.324±0.020	0.532±0.051	0.331±0.054	0.315±0.039	0.420±0.027	0.450±0.011	0.421±0.091
	Zeta-potential (mV)	- 27.2 ±0.4	- 28.9± 0.5	- 28.6±1.5	- 31.6 ±2.2	- 28.9±0.2	- 32.7± 0.1	- 33.5±3.2	- 34.0±3.3	- 45.1. ±2.1	- 36.2±7.1
4 weeks	Size (nm)	186.8±1.6	182.0±2.7	311.8±8.1	190.0±2.5	255.8±34.8	201.2±4.0	183.5±9.1	260.6±26.4	221.2±12.2	210.3±5.1
	PDI	0.138±0.025	0.226±0.016	0.364±0.042	0.208±0.037	0.534±0.078	0.340±0.019	0.310±0.011	0.420±0.027	0.450±0.011	0.421±0.091
	Zeta-potential (mV)	- 25.5 ±1.3	- 27.3± 1.5	- 26.7±0.2	- 27.8 ±1.0	- 29.8±1.1	- 32.5± 0.1	- 33.2±1.7	- 34.0±3.3	- 45.1±2.1	- 36.2±7.1
Fresh samples		P1	P2	P3	P4	P5	DP1	DP2	DP3	DP4	DP5
	Size (nm)	195.3±21.5	178.1±3.0	169.3±19.2	180.6±7.2	173.5±42.6	176.7±11.4	156.6±10.2	147.6±14.7	168.2±12.0	137.1±4.1
	PDI	0.462±0.055	0.417±0.027	0.435±0.098	0.447±0.076	0.383±0.025	0.493±0.010	0.450±0.009	0.448±0.095	0.481±0.072	0.437±0.046
1 week	Zeta-potential (mV)	- 35.9± 2.0	- 43.8± 0.6	- 49.1± 1.3	- 38.9±3.2	- 47.0± 1.4	- 27.7±2.0	- 26.5± 0.8	- 32.1± 1.0	- 39.0 ±2.4	- 41.8±2.7
	Size (nm)	193.2±11.1	175.2±5.6	165.3±12.1	181.5±5.2	170.5±22.7	171.7±10.1	157.2±10.3	146.3±11.6	167.5±12.8	141.1±4.1
	PDI	0.342±0.029	0.317±0.011	0.397±0.018	0.417±0.016	0.313±0.011	0.393±0.032	0.390±0.019	0.380±0.095	0.430±0.011	0.430±0.146
2 weeks	Zeta-potential (mV)	- 32.2± 3.0	- 39.1± 0.6	- 45.1± 7.3	- 35.3±1.2	- 45.0± 1.3	- 27.5±1.4	- 25.5± 0.4	- 31.2± 0.6	- 35.1 ±2.1	- 39.2±5.7
	Size (nm)	195.0±10.2	172.3±5.2	170.3±22.2	186.5±10.1	175.9±12.2	170.7±10.2	155.0±10.4	149.4±15.2	168.9±21.1	144.6±7.2
	PDI	0.331±0.039	0.310±0.011	0.410±1.018	0.419±0.026	0.321±0.021	0.389±0.011	0.379±0.013	0.389±0.011	0.410±0.025	0.429±0.210
3 weeks	Zeta-potential (mV)	- 32.0± 3.7	- 39.28± 0.2	- 45.1± 4.3	- 35.6±2.1	- 43.6± 1.5	- 27.2±1.1	- 26.1± 2.0	- 31.2± 0.1	- 35.9 ±2.2	- 39.5±3.5
	Size (nm)	191.1±10.2	175.9±5.1	198.3±22.2	195.5±8.2	189.9±10.2	172.3±15.1	154.3±10.2	170.2±10.4	192.3±19.3	177.2±7.1
	PDI	0.310±0.039	0.335±0.011	0.397±1.018	0.410±0.026	0.325±0.021	0.390±0.012	0.389±0.313	0.391±0.321	0.419±0.011	0.437±0.217
4 weeks	Zeta-potential (mV)	- 32.9± 4.0	- 39.3± 0.5	- 45.6± 4.3	- 35.2±2.3	- 44.9± 1.4	- 27.9±3.1	- 26.5± 3.4	- 31.4± 1.2	- 35.5 ±2.3	- 39.3±4.2
	Size (nm)	194.0±10.1	176.4±5.3	207.3±22.1	210.5±8.1	197.9±10.2	171.7±15.1	156.4±9.6	183.4±15.1	212.4±21.1	191.6±7.3
	PDI	0.350±0.039	0.310±0.011	0.381±1.018	0.410±0.026	0.327±0.021	0.370±0.012	0.376±0.213	0.389±0.011	0.431±0.025	0.423±0.210
	Zeta-potential (mV)	- 32.3± 4.2	- 39.3± 0.6	- 45.9± 4.2	- 35.2±2.2	- 43.5±1.4	- 27.4±3.1	- 26.9± 2.1	- 31.2± 0.2	- 35.2 ±2.3	- 39.7±3.6

3.5 Interaction studies

3.5.1 ATR-FTIR spectroscopy

The ATR-FTIR spectroscopy is a pre-formulation study for the evaluation of compatibility between the formulation ingredients. The ATR-FTIR spectra of DOX, PXT, and all the other ingredients individually, the physical mixtures of formulations, and their corresponding niosomal formulations are shown in Figure 2.

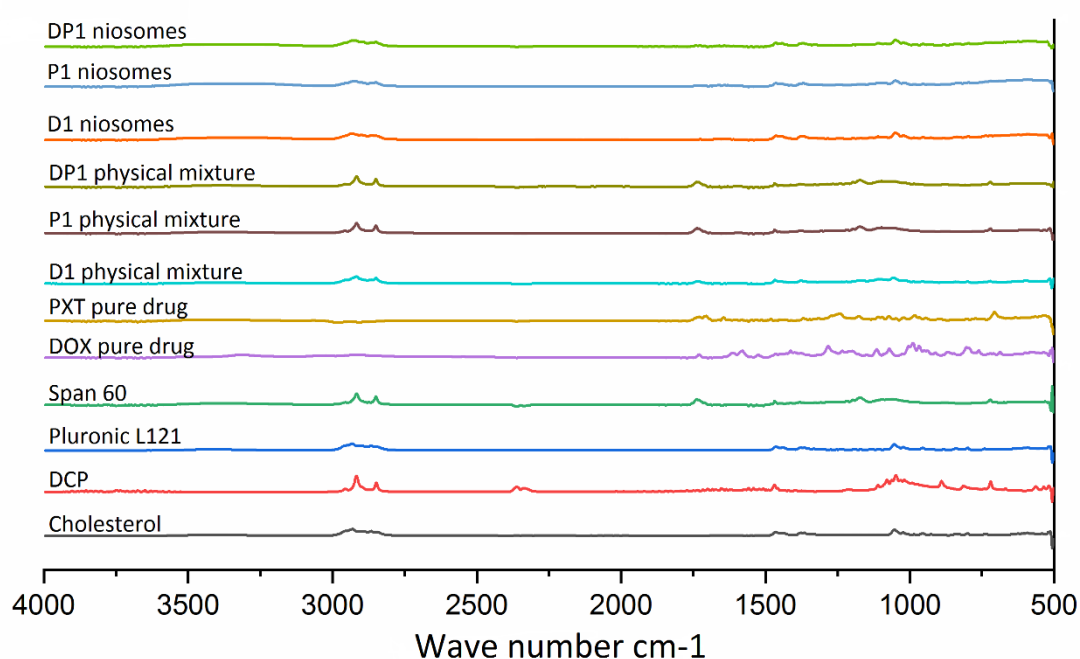


Figure 2. ATR-FTIR spectra of different raw materials, physical mixtures and niosomal formulations.

The pure DOX showed peaks at 3456 cm^{-1} and 3335 cm^{-1} due to the N-H stretching of primary amine and O-H stretching, respectively. The peaks at 868 cm^{-1} and 807 cm^{-1} were seen due to N-H group stretching (Majeed *et al.*, 2013). The ATR-FTIR spectrum of PXT (pure drug) showed peaks between 3479 cm^{-1} to 3300 cm^{-1} due to the stretching of N-H, CH_2 stretching peaks between $2976\text{--}2885\text{ cm}^{-1}$, C=O stretching at 1734 cm^{-1} , the amide bond stretching at 1647 cm^{-1} , ester bond and C-N stretching at 1254 cm^{-1} and 1276 cm^{-1} ,

respectively, and peaks due to aromatic bonds were seen at 1647, 1074, 963 and 709 cm^{-1} (Martins *et al.*, 2014). Span 60 gave peaks at 2916.75 cm^{-1} due to (-OH stretch, broad), 2849.58 cm^{-1} (-OH stretch, broad), a 5-membered cyclic ring peak at 1734.65 cm^{-1} and small peaks due to aliphatic groups from 1000-1200 cm^{-1} (Li *et al.*, 2008) Pluronic L121 showed peak stretch at 2990 cm^{-1} of asymmetrical methyl (C-H), scissoring of C-H bondage at 1480 cm^{-1} , symmetrical C-H bond at 1387 cm^{-1} and ether linkage (C-O-C) at 1120 cm^{-1} (Newman *et al.*, 1998). Cholesterol showed ATR-FTIR peaks at 2931.41 cm^{-1} of acetyl group, 2866.83 cm^{-1} symmetric -CH₃, at 1770.20 cm^{-1} and 1055.17 cm^{-1} due to vinyl group and R-O groups, respectively (Khan *et al.*, 2015).

The ATR-FTIR of the physical mixtures of the optimized formulations and their relevant niosomal formulations were also studied. The spectra of both the physical mixtures and the niosomal formulations were similar and diffusion of the peaks were seen without peak shifting. Similar interactions between the ingredients was seen as in the earlier studies, where it was suggested that there was interactions between the Span 60 and cholesterol to give rigid and stable structure for niosomes (Nasseri, 2005). The found interaction is between the glycerol oxygen in Span molecule and β -OH group in cholesterol molecule; changes due to this interaction were seen in the spectrograms.

3.5.2 Thermal analysis

In the thermal analysis of Span 60, DCP, cholesterol, DOX and PXT, they showed endothermic characteristic melting peaks at 54°C, 78°C, 150°C, 205°C and 220°C, respectively (Figure 3). The physical mixtures of formulations (D1, P1 and DP1) showed the slightly broader peaks at 38°C and 57°C which indicates the interaction between the Span 60 and cholesterol (Nasseri, 2005), as already mentioned related to the discussion of ATR-FTIR results, and which was also seen with the niosomal formulations D1, P1 and

DP1. Also, a slightly broader peak was seen at 140 °C due to cholesterol. The peaks of membrane formers indicated mutual interactions, which resulted in more stable niosomes as indicated by the stability studies data and also by the entrapment efficiency values. The slightly broader peaks of drugs were seen in the range of 205°C to 230°C which indicated that the drugs were in their crystalline form as have been indicated in previous studies (Pawar and Vavia, 2016; Doustgani, 2017).

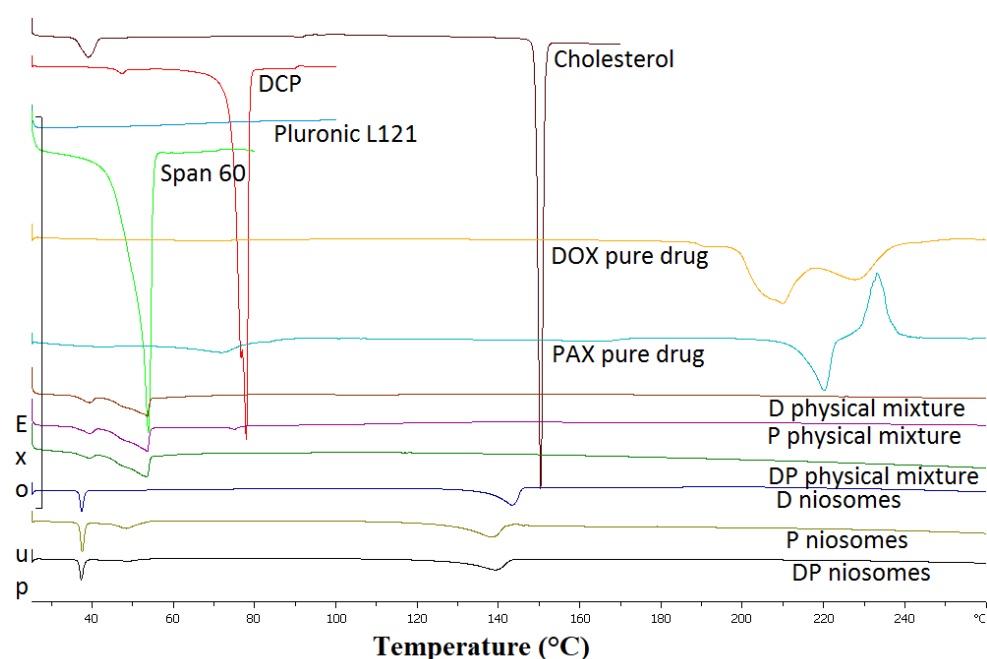


Figure 3. DSC thermograms of pure materials, physical mixtures and niosome formulations with PAX (P), DOX (D) and DOX and PAX (DP).

3.6 Drug Release Studies

The *in vitro* drug release testing from the niosomes was performed in HBSS-HEPES (pH 7.4) with 4% Tween 80 addition (Yang *et al.*, 2007a; Figueiredo *et al.*, 2017). The release profiles of DOX and PXT are summarized in Figure 4. From pure DOX powder, the dissolution was very fast. Drug release from the DOX niosomes was biphasic in nature: in the first phase, burst type of release was seen, which was followed by a constant sustained release phase. Dissolution rate from DOX niosomes was a little bit slower as compared to pure drug: from niosome formulations containing only DOX, drug release after 24 h time was approximately 93%. Also, with the formulations containing both DOX and PXT, same kind of burst release was seen, but it was comparatively in lower level as compared to the formulations containing only DOX. Later, a constant sustained release of DOX from DOX and PXT loaded niosomes was observed reaching approximately 40% level within 24 h.

Also with PXT, the biphasic release was observed. In the first phase, burst type release was seen and later a constant sustained release of PXT was seen from formulations containing only PXT reaching the approximated level of 26% within 24 h. Again, with the formulations containing both DOX and PXT, a burst release of PXT was seen, but similarly to DOX release, it was less as compared to the formulations containing only PXT. In the second phase, a constant sustained release of PXT was observed reaching the approximated level of 18% within 24 h. All the PXT loaded niosomes showed much higher dissolution rates as compared to pure PXT.

The niosomal formulations containing both DOX and PXT showed lower drug release rates as compared with the formulations containing only one drug.

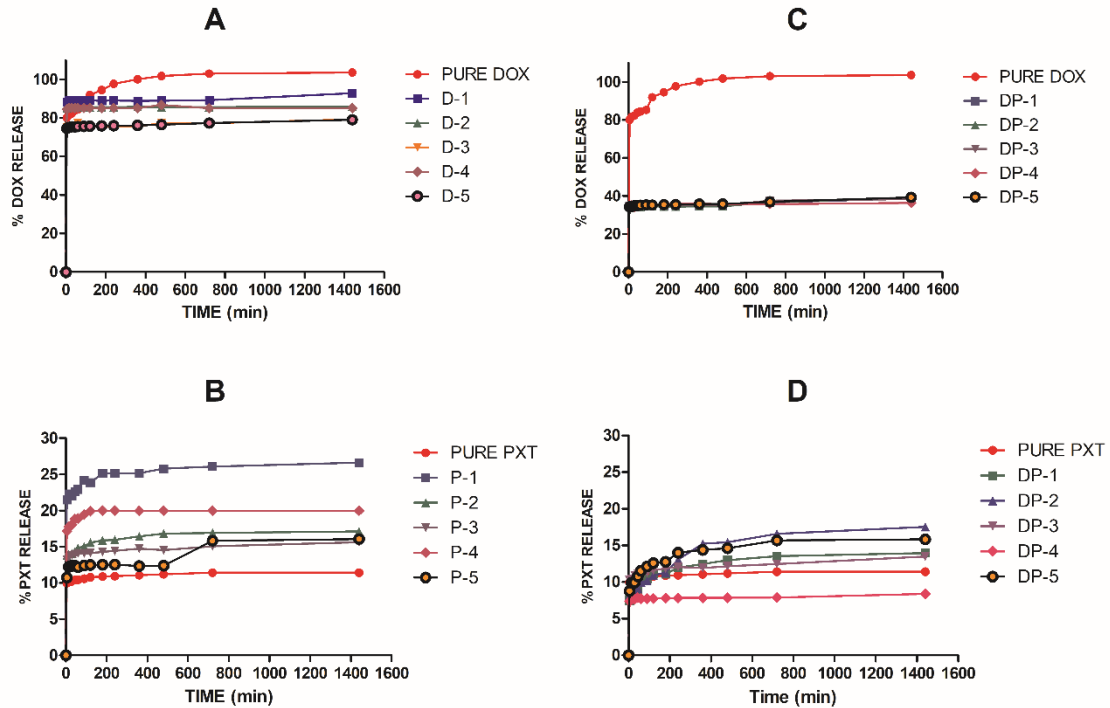


Figure 4. Drug release from formulations containing A) DOX, B) PXT, and formulations containing DOX and PXT, release of C) DOX, and D) PXT.

3.7 Cell viability assay

The biocompatibility of the niosomes is an important criterion for their application as a drug delivery system. In this study, the cytotoxicity of the niosomes was evaluated against MCF-7 and PC3-MM2 cell lines by incubating them for 24 h using different niosome concentrations as shown in Figure 5. With empty niosomes, weak concentration related effect on viability was noticed as compared to the negative control, but the viability of the cells with even the highest tested niosome concentration, 1 000 $\mu\text{g/ml}$ of niosomes, was not significantly lowered with either MCF-7 or PC3-MM2 cell lines (with both cell lines $p > 0.05$). With all the studied concentrations, the viability values were higher with MCF-7 cells as compared to PC3-MM2 cells.

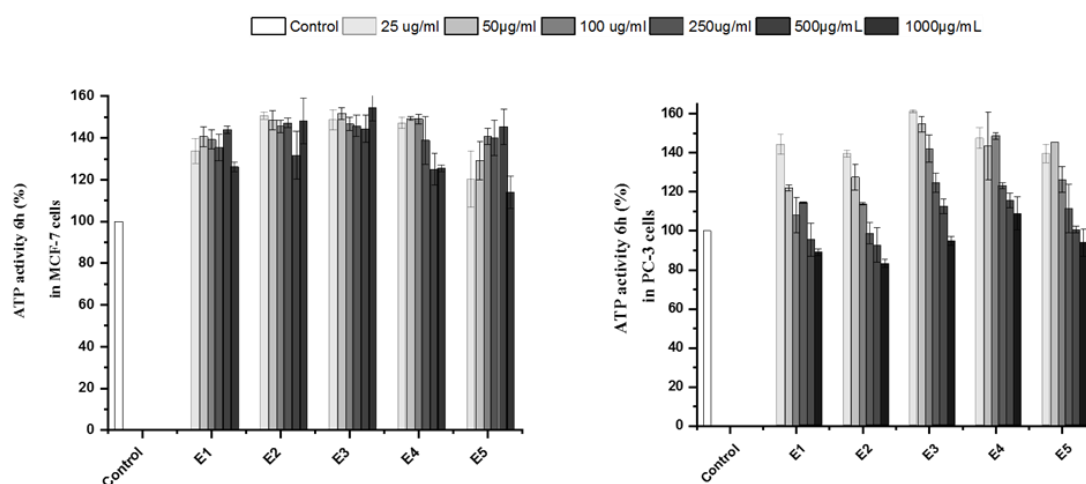


Figure 5. Cell viability assay of niosomal formulations against MCF-7 and PC3-MM2 cells after 24 h incubation.

3.8 Cell uptake studies

The cell uptake efficiency of the anticancer drug loaded niosomes is an important factor for the estimation of potency of drug formulations. A fluorescent compound, FITC, was incorporated into the niosomes, cell membranes were stained with CellMask Deep Red and nuclei with DAPI. The confocal images (Figure 6) of MCF-7 and PC3-MM2 cell lines were examined by incubating them at 37°C for 6 h. Based on the confocal images, the niosomes were taken up by the cells and they were accumulated into the cytoplasm of the cells successfully. In earlier studies, it has been shown that the internalization of niosomes can be facilitated by the presence of lipid content of niosomes and its interaction with the lipid membrane of the cells (Guo *et al.*, 2015; Shi *et al.*, 2015; Tahir *et al.*, 2017). The size of the niosomes also affects internalization and drug delivery into the cells. In the present study, the smallest size fractions (<200 nm) of niosomes had better penetration due to having small size, and the larger niosomes had less penetration capacity, which has also been described in previous studies (Shahbazi *et al.*, 2014; Khutale and Casey, 2017). The formulation E2 showed the best internalization of the niosomes when compared with other

formulations. In other formulations, part of the niosomes remained outside of the cells and adhered to cell surfaces.

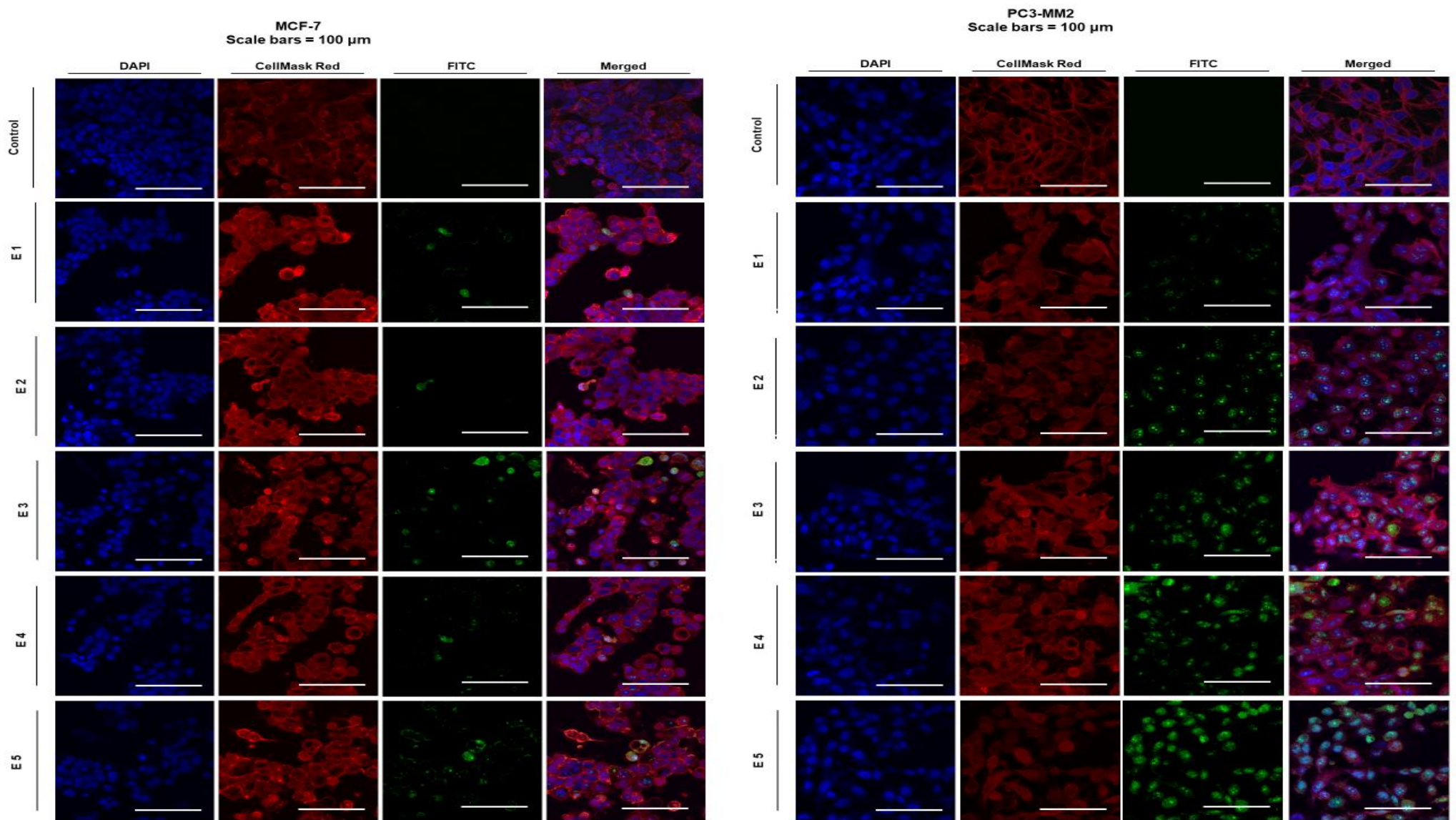


Figure 6. Confocal images of niosomal formulations uptake by MCF-7 and PC3-MM2 cells.

3.9 Antiproliferative studies

The antiproliferative activity of the anticancer drugs (DOX, PXT) loaded niosomes was measured by ATP activity based luminescence assay as indicated in Figure 7. The effect of pure drugs, empty niosomes, and niosomes containing the anticancer drugs, was evaluated on MCF-7 and PC3-MM2 cell lines. The effect of the pure drugs was seen to be concentration dependent. DOX showed more antiproliferative effect on PC3-MM2 cells as compared to PXT pure drug at highest concentration levels (500 µg/ml, $P<0.05$), but in MCF-7 cell line there were no statistical difference in antiproliferative effect between the pure drugs in the highest concentration. The niosomal formulations showed time and concentration dependent type of activity.

In MCF-7 cell line the formulations containing both drugs didn't differed from formulations having either of the drug alone. But, In PC3-MM2 cells, dual drug therapy (DOX+PTX niosomes) showed better antiproliferative effect on cancerous cell lines and efficacy of the niosomal formulations as compared to either DOX alone ($P<0.01$) or PTX alone ($P<0.05$). The formulations containing paclitaxel and doxorubicin (DP1-DP5) showed improvement of antiproliferation against MCF-7 and PC3-MM2 cells. The efficacy against PC3-MM2 is much higher than MCF-7 ($P<0.01$). It showed concentration and time dependent activity.

The *in-vitro* release studies showed the sustained drug release behavior of the formulations, which is also clear from the cell line antiproliferation studies. The increase in incubation time of cells would improve the effects. But, the results were in line with the results of release studies of the formulations, which was sustained drug release which is also reported in previous findings (Tahir *et al.*, 2017; Alemi *et al.*, 2018).

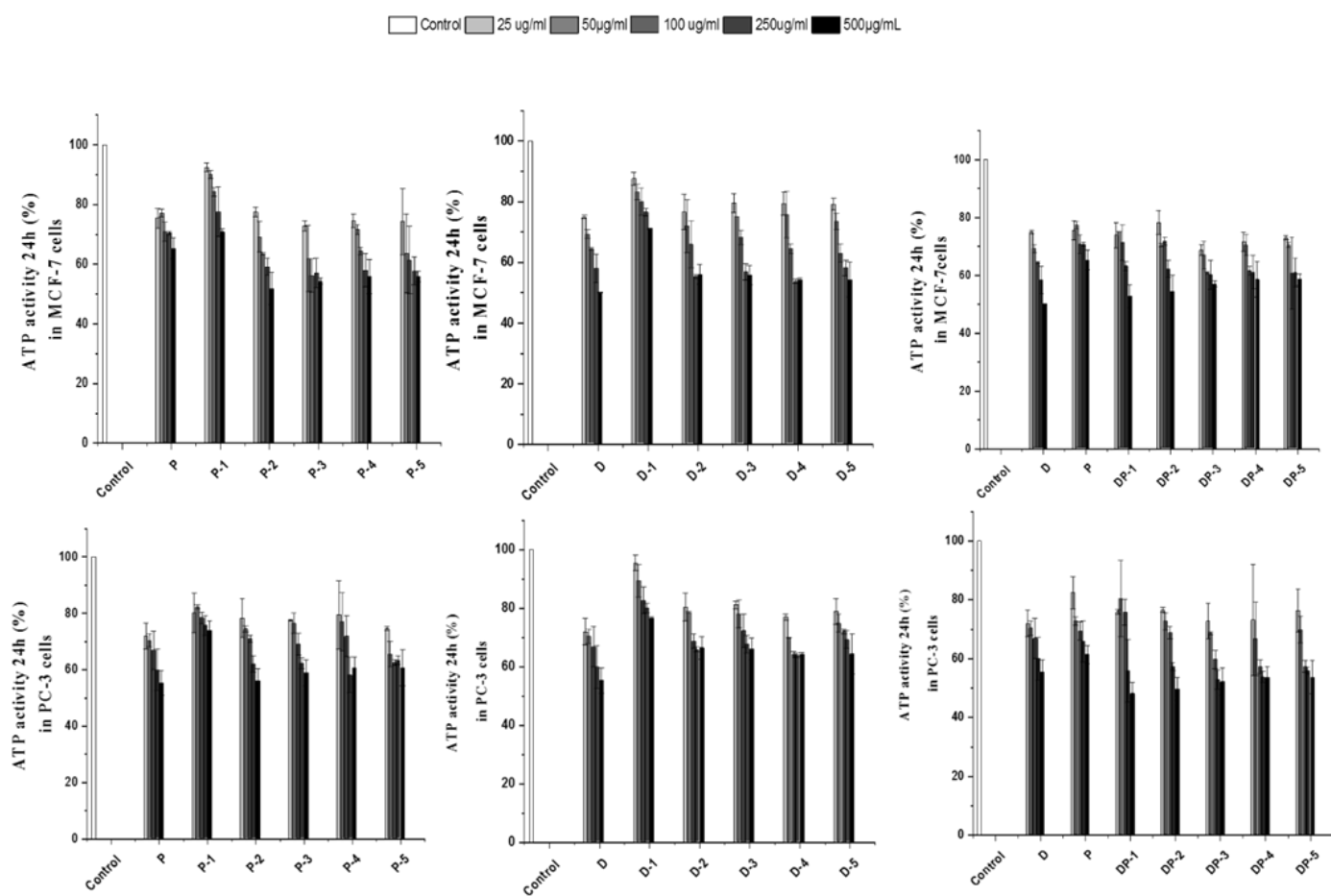


Figure 7. Antiproliferative study of different concentrations of drugs on MCF-7 and PC3-MM2 cell after 24 h incubation time.

Conclusion

In the present study, two cytotoxic drugs, doxorubicin and paclitaxel, were encapsulated into niosome formulations by using an ecofriendly probe sonication method. The niosomes prepared with fixed amount of Span 60 and cholesterol, and with varying amount of Pluronic L121 and dicetylphosphate showed high entrapment efficiencies, as well as acceptable size and monodispersity levels. *In-vitro* characterization showed that the formulations had sustained drug release profiles, low toxicity even at high concentrations, better cell penetration and improved antiproliferative effects in time and dose dependent manner. Further, niosomes showed synergistic effect due to the presence of two anticancer drugs.

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